



## Antifungal Activity of Derum (Juglans Regia L. Bark) Extracts Against Candida Albicans Isolates (In Vitro Study)

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### Abstract

**Background:** Derum is a one of "chewing sticks" existing in nature, collected from stem bark of walnut tree (*Juglan regia L.*). In some countries used mainly by women as tools for cleaning teeth and as coloring material for cosmetic purpose.

The objective of performed study was aimed at evaluating the antifungal potential of two extracts from *Juglans regia (L.)* bark (water and water-methanol) against pathogenic *Candida albicans* strains.

**Materials and methods:** Different concentrations of derum extracts were prepared by (water and water-methanol) methods, swap of saliva were collected from volunteers of patient at dental hospital, from which *Candida albicans* were isolated, purified and diagnosis according to morphological characteristic and biochemical test. In vitro; experiments were prepared to assess the effect of different type and concentrations of derum extracts on growth of *Candida albicans* by agar diffusion methods in comparison with nystatin and amphotericin B.

**Results:** The result showed, *Candida albicans* isolates sensitive to both types of derum extracts, which improved, by inhibition growth of *Candida albicans* and inhibition were increased with the increasing concentrations from 0.5% to 50%. Nystatin inhibition growth of *Candida albicans* better than both types of derum extracts (water and alcohol) and amphotericin B; but *Candida albicans* more sensitive to both types of derum extracts than amphotericin B.

**Conclusion:** *Candida albicans* were sensitive to different concentrations of (water and water-methanol) extracts of derum.

**Key words, Candida albicans, Juglans regia bark extracts, Antifungal activity.**

### Introduction

"For many centuries different populations and cultures around world have been using various tools, ranging from porcupine bones to chewing sticks to clean their teeth and gum"<sup>(1,2)</sup>. Relatively easy to access and the popularity of "chewing sticks", that have made a different communities were used it as tools for plaque control; although "chewing sticks" obtained

from different sources, many components of chewing sticks and other related plants were reported to have beneficial biological properties, including significant anti-bacterial and anti-fungal activity<sup>(3,4)</sup>. Derum is a one of "chewing sticks" existing in nature, collected from stem bark of walnut tree (*Juglan regia L.*). In some countries used mainly by women as tools for

cleaning teeth and as coloring material for cosmetic purpose<sup>(5)</sup>. Its extracts showed broad-spectrum antimicrobial activity. It inhibition microorganisms including gram-positive bacteria like (*Staphylococcus aureus* and *Streptococcus mutans*), gram-negative bacteria like (*Escherichia coli* and *Pseudomonas aeruginosa*) and different types of pathogenic yeast for example (*Candida albicans*); derum extract also increase the pH of the saliva<sup>(6,7)</sup>. Different extract of *juglan regia* L. bark show potent antioxidant activity<sup>(8)</sup>.

"*Candida* is a genus of yeast, commonly, it is part of the normal flora of the mouth, skin, intestinal tract and vagina"<sup>(9,10)</sup>. But sometimes in individuals with predisposing factors, which present among immunodeficiency, diabetic, age extreme radiotherapy, AIDS, treated with antibiotic and pregnant women, denture wearing and the use of orthodontic appliance, candidiasis is the earliest infection manifest for million people<sup>(11,12)</sup>. *Candida albicans* strains consider the most commonly species that causing invasive candidiasis of skin surface, mucosal membrane, urinary tract infection and even life threatening by systemic invasions with candidaemia<sup>(13)</sup>. The most common treatment is antifungal agents, such as (fluconazole, itraconazol, miconazol and ketoconazol) and polyenes (amphotericin B or nystatin)<sup>(14)</sup>. Unfortunately, antifungal therapy is very limited in addition to prolong uses of antifungal treatment will produce problems of side effects for example hematological, hepatic, and/or renal toxicity and may be lead to emergence of strain resistant<sup>(15)</sup>. The challenge has been to develop effective strategies for treatment of candidiasis and other fungal disease. Lower side effects of very types of herbal extracts have

suggested them as sources of new pharmaceuticals<sup>(16)</sup>. Recently, many plant extracts have therapeutic value with respect to oral diseases<sup>(17)</sup>.

The present study performed to test the effect of different concentrations of derum extracts (water and alcohol) on growth of *Candida albicans* in comparison with other two anti-fungal therapy which are nystatin and amphotericine B.

## Materials and methods

### Preparation of derum extracts:

The stem bark of *juglan regia* L. obtained from local market. Derum sticks were cut into small pieces and crunching to powder using a roller-mill. water and water-methanol extracts was prepared by adding 100g of small partials from plant material to 500ml of solvent (deionized water and methanol 99.8%) and allowing the mixture to stand 48 hours at room temperature<sup>(18)</sup>. Then it was filtered by using filter paper (No. 1). The solvents were evaporated at 40°C by means of rotary evaporator to get crud extracts, then the both extracts dissolved in water to prepare different concentrations.

### Fungal isolates:

Isolates of *Candida albicans* strain had been obtained from oral cavity of patients with orthodontic appliance in dental hospital by swabbing method; no patient was under antifungal drugs. A sterile cotton swab (NipponMenbo, Tokyo, Japan) was immediately cultured into sabouraud dextrose agar (Hi Medic, Mumbai, India) to elicit isolated; which incubated at 37°C for 24-48 hour. Colonies of *Candida albicans* strain purified and diagnosis according to morphological characteristic and biochemical test.

The anticandidal potential of investigated extracts was screened by using disk diffusion method of Noumi et al with minor modifications<sup>(19)</sup>; according to Noumi et al inoculum was prepared by adding colonies of candida albicans strain to normal saline 0.85% and adjusted to McFarland 0.5 turbidity standard which contain  $10^6$  cell/ml. Adequate amount of Mueller Hinton agar (Hi-Media) (with 2% glucose were distributed into sterile plates and allow solidifying under aseptic condition. The organism suspension was inoculated with sterile spreader on surface of solid medium in plates. Four wells of equal size and depth had been prepared in each agar plate; each well was filled with 0.1 ml of the test agents. The plates were incubated for at 37°C for 24 h. Zone of inhibition which is clear zone of no growth were measure across the diameter of each well. No zone indicated complete resistance of fungi to the agents. As control positive nystatin 100,000 IU/ml ( Lifepharmaco, Italfarmaco, Group - Italy ) and amphotericin B 10 µg/ml (Fungizone, BioBasic, Inc., Toronto, Canda) were used.

### Statistical Analysis:

Data manipulating and anaylasis were done by using SPSS program version 21.

### Result

Sensitivities of candid albicans to different concentrations of derum extracts (water and alcohol), nystatin and amphotericin B were tested according to Agar Well Technique. The results showed that both type of derum extracts had same color dark brown as showed at (**figure1**). Candida albicans were sensitive to water and water-methanol extracts of derum by inhibition growth of candida albicans

strain and the sensitivity increased with increasing concentrations start from (0.5%) to (50%) which more effective, candida albicans were more sensitive to nystatin compare to both types of derum extracts and amphotricien B as showed at (**figure2**).

**Table (1)**, showed mean and Standard Deviation(SD) for sensitivity of candida albicans to different agents.

**Table (2)**, showed comparison between both type of derum extracts (water and methanol) at same concentration, statistically no significant difference for inhibition growth of candida albicans between 0.5% and 1% concentration and high significant difference between 5%,10%,15%,and 20%, but a significant difference was found between 30%,40% and 50%.

**Table (3)**, preformed to comper each concentration of either water or water methanol extracts with nystatin and amphotricine B, for water extract, candid albicans sensitive to nystatin higher than water extracts, statically with a highly significant difference. The result showed that candida albicans sensitive to water extracts higher than amphtricin B at 40%and 50% with highly significant difference. On comparison water-methanol extracts with nystatin, candida albicans sensitive to nystatin more than water- methanol extract with highly significant difference from 0.5% to 30% concentrations and a significant difference at 40% and 50% but when comparsion with amphotericin B, the result showed that water methanol extract better than amphotricic B for inhibition growth of candida albicans with highly significant difference from

5% to 50% with no significant difference at concentration 0.5% and 1%. The result showed that nystatin better than amphotericin B for inhibition growth of *Candida albicans* with highly significant difference.

## Discussion

A variety of chemotherapeutic agents have been investigated for a possible ability to control oral pathogens. Herbal materials are one of groups which have been studied in such investigations. The present study was designed to obtain information on the microbial effect of juglan regia bark on *Candida albicans* and findings have validated the presence of such effect. At present study evidence support an antifungal effect of derum on *Candida albicans*. Polyphenol and derivatives are not only antimicrobial compounds isolated from juglan regia bark<sup>(5,7)</sup>, but juglon and glycosides are amongst strongest antimicrobials constituents of juglan regia bark<sup>(20,21)</sup>. The anti-microbial effects of derum was observed on gram-positive, gram-negative and fungi<sup>(6,7)</sup>. In spite of, there is a drib information exist about antifungal attributes of derum spatially against *Candida albicans* some study report that (water and alcoholic) extracts of juglan regia were inhibited in vitro growth, *Candida albicans* strains<sup>(7,16,19,22)</sup>, and that coincidence with this study. The antifungal effect of derum against *Candida* strain may be through the following mechanisms; The number of hydroxyl group (OH)<sup>-</sup> on the some constituents of derum specially for the polyphenol were believed to be related to their relative toxicity to microorganisms, with evidence that increased hydroxylation result in increased toxicity<sup>(23)</sup>. Now there are many of researches about using phenolic substance in order to inhibit

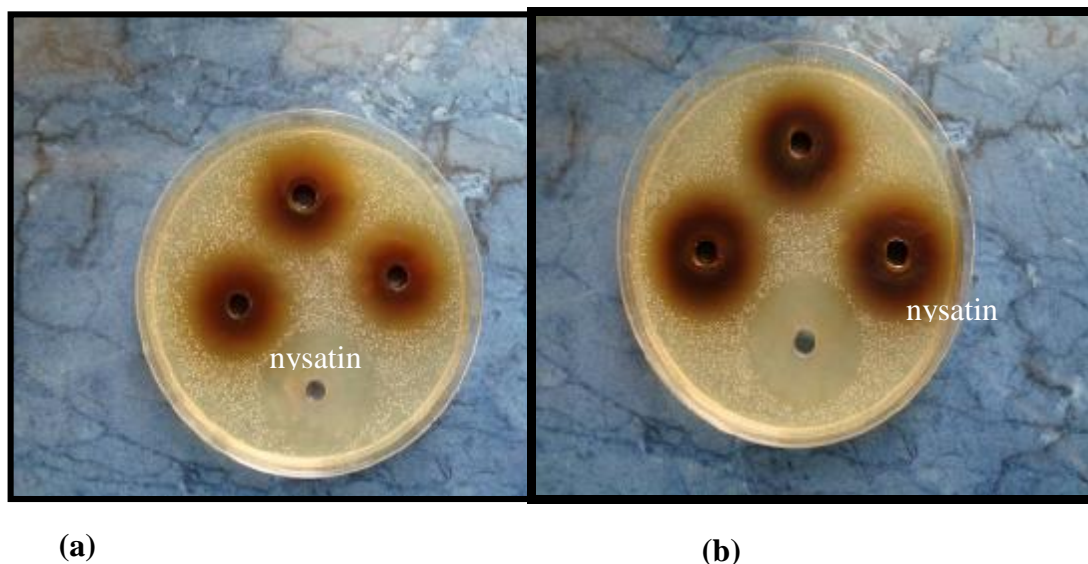
the growth of fungi<sup>(24, 25)</sup>, as we know the main *Candida* virulence factors are exoenzymes production, biofilm-formation, adherence and dimorphism<sup>(26,27,28)</sup>; many studies improved the effect of phenolic acids against these factors<sup>(29,30,31)</sup>. Another mechanism is by the inhibition of biofilm formation of *Candida albicans*; once this biofilm is inhibited, the fungi cell will lose their connection to each other and the surface which apparently can cause cell death<sup>(32,33)</sup>. Juglon has believed to inhibit broad spectrum of microorganism including bacteria, and fungi including *Candida albicans*. Many recent studies have focused on the toxicity effect of juglon toward microorganisms<sup>(1)</sup>. The mechanisms for the toxic effects of juglon is not clearly understood, several pathway have been proposed which include that it may be cause death of cell, disruption cell cycle, modification of DNA, inhibition synthesis of mRNA, alkylation of thiol or amine groups of essential protein, and decreasing level of p53 (tumour suppressor)<sup>(34,35)</sup>. Juglon can also be reducing enzyme in mitochondria or cytoplasm to form a semi-quinone radical<sup>(36)</sup>.

## References

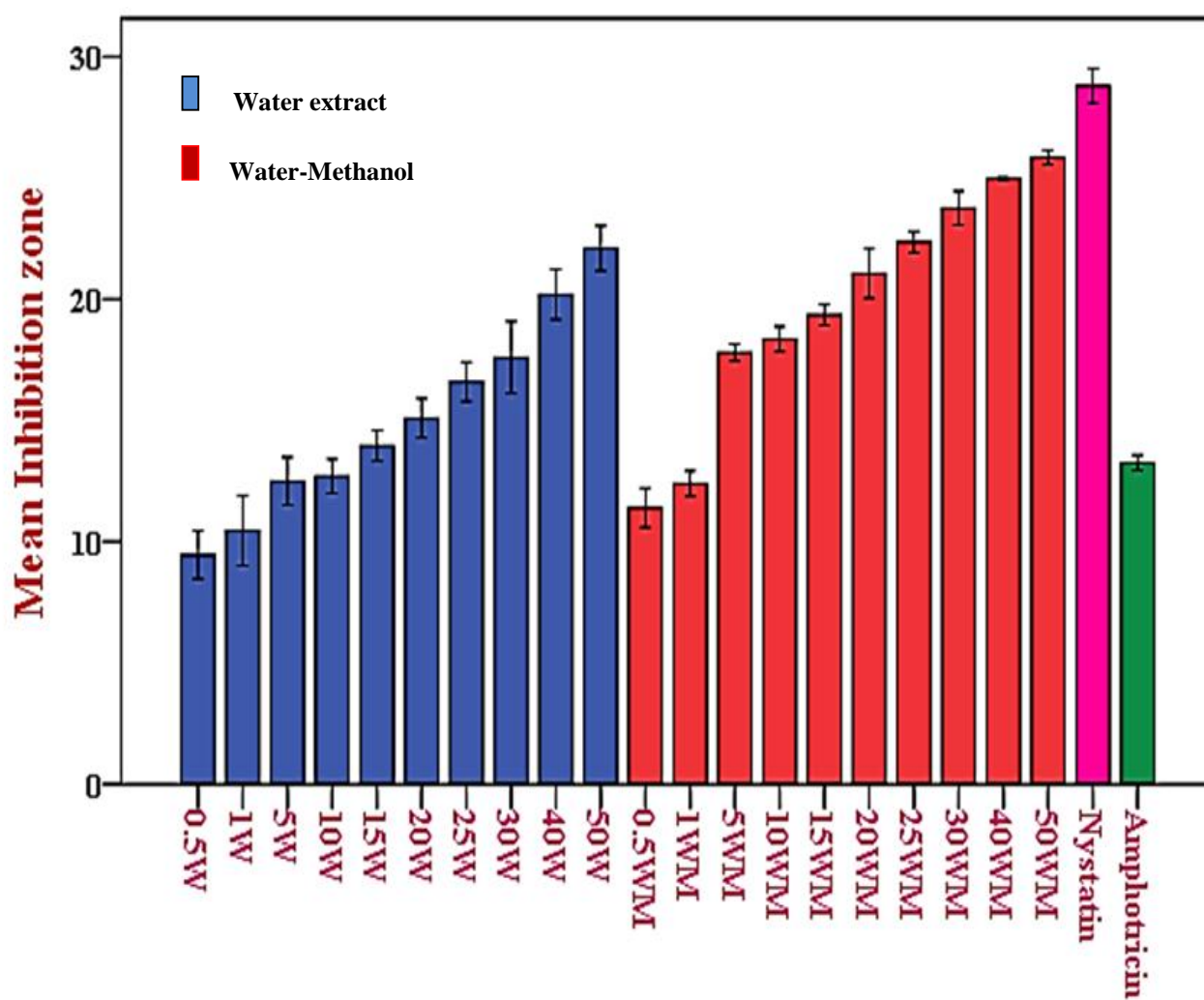
- 1- Khoory T. The use of chewing sticks in preventive oral hygiene. *Clin Rev Dent*. 1983: 11-14.
- 2- Wu CD., Darout IA., Skang N. Chewing sticks: timeless natural tooth brushes for oral cleaning. *J. Periodontal Res*. 2001; 36: 275-284.
- 3- Pulger B., Gonuz A. Antimicrobial activity of certain plant used in Turkish traditional medicine. *Asian J. Plant*. 2004; 3: 104-107.
- 4- Brchord R., Donald L., Kendral S., Fulcher R., Bey R. Antimicrobial activity of nature and naturalize plant of Minnesota and wisconsin. *Journal of Medicinal Plant Research*. 2008; 2(5): 98-110.
- 5- Osman NA., Gafar SM., Salah el-Din M., Wassel GM., Ammar NM. Hazardous effect of topical cosmetic application of

- deirum(*juglan regia* plant) on oral tissue. *Egypt Dent J.* 1987 Jan; 33(1): 31-35.
- 6- JagtapAG., Karkera SG. Extract of *juglandaceae regia* on growth, in-vitro adherence, acid production and aggregation of *streptococcus mutans*. *J Pharm Pharmacol.* 2000 Feb; 52(2):235-42.
  - 7- Rahul R. et al. Antimicrobial activity of different extracts of *juglan regia* L. against oral micro flora. *International journal of Pharmacy and Pharmaceutical Science.* 2011; 3(2): 200-201.
  - 8- Agarwal K. and Chakarborthy GS. In vitro antioxidant activity of different extracts of bark of *juglan regia*. *International journal of Innovative Pharmaceutical Research.* 2013;3(1): 199-202.
  - 9- Paluchowska P., Tokarczyk M. Boguzz B., Skiba I., Budak A. molecular epidemiology of *candida albicans* and *candida glabrata* strains isolated from intensive care unit patients in poland. *Mem.inst. Oswaldo Gruz.* 2014 Jul; 109(4): 436-41.
  - 10- Hube B. From commensal to pathogen: stage- and tissue-specific gene expression of *Candida albicans*. *Current Opinion in Microbiology.* 2004; 7(4): 336-341.
  - 11- Hamza OJ. et al. Species distribution and in vitro antifungal susceptibility of oral yeast isolates from Tanzanian HIV-infected patients with primary and recurrent oropharyngeal candidiasis. *BMC Microbiology.*2008; 8:135
  - 12- Kumar BV., Padshetty NS., Bai KY. And Rao MS. Prevalence of *candida* in the oral cavity in diabetic subjects. *J. Assoc. Physician. India.* 2005; 53: 599-602.
  - 13- Pfaller MA., Diekema DJ. Epidemiology of invasive candidiasis a persistent public health problem. *Clin Microbial Rev.* 2007; 20: 133-63.
  - 14- BurketLW., GreenbergMS. GlickM. Ship JA. *Burket's Oral Medicine: Diagnosis & Treatment.* Eleventh ed. Hamilton Ontario. Bc. Decker Inc., 2008:39.
  - 15- Hope WW. et al. Molecular Mechanisms of Primary Resistance to Flucytosine in *Candida albicans*. *Antimicrob Agents Chemother.* 2004 Nov; 48(11): 4377-4386.
  - 16- Abad MJ., Ansuategui M., and Bermejo P. Active antifungal substances from natural sources. *ARKIVOC.* 2007; (vii): 116-145.
  - 17- Sharma S., Vijayvergiu R., Sinsh T. Evaluation of antimicrobial efficacy of some medical plants. *J. Chem. Pharm. Res.,* 2010;2(1): 121-124.
  - 18- Moori BN., Khalafi E. Anti-bacterial activity of hydro-alcohol extracts on *juglan regia* L. stem bark on human bacterial infection. *Quarterly of international Archives of health sciences.* 2015; 2(4): 139-143.
  - 19- Noumi E., Snoussi N., Hajlaoui H., Valenten E. and Bakhrouf A. Antifungal properties of *salvadora percia* and *juglan regia* L. extracts against *candida* strains. *European.Journa of clinical Microbial. Infectious Diseases.* 2010; 29: 81.
  - 20- Clark AM., Jurgens TM., Hufford CD. Antimicrobial activity of *juglon*. *Phytotherapy Research.* 1990; 4(1): 11-14.
  - 21- Poyla GM. Biochemical targets of plant bioactive compounds. First ed. CRC/Press, 2003: 384.
  - 22- Sytykiewicz H. et al. Antifungal activity of *juglan regia* L. leaf extracts against *candida albicans* isolates. *Pol. J. Environment Stud.* 2015; 24(3):1339-1348.
  - 23- Geissman TA. Flavonoid compounds, tannis, lignins and related compounds. In: Florkin M and Stotz EH (ed.). *Pyrol pigments, isoprenoid compounds and phenolic plant constituents.* Elsevier, New York, N.Y. (cited).1963; 9: 265
  - 24- Perira JA., et al. Bioactive properties and chemical composition of walnut(*juglan regia* L.) selection from eastern turkey. *Afr. J. Agric. Res.,* 2010; 5:2379-2385.
  - 25- Oliveria I., et al. Total phenols, antioxidants potential and antimicrobial activity of walnut(*juglan regia* L.) green husks. *Food Chem. Toxicol.* 2008; 46: 2326-2331.
  - 26- Vuong C., Kocianova S., Voyich JM., Yao Y., Fischer ER., Deleo FR., et al. A crucial role for exopolysaccharide modification in bacterial biofilm formation, immune evasion, and virulence. *J. Biol. Chem.*2004; 279:54881-54886.
  - 27- Netea MG., Brown GD., Kullberg BJ., Gow NA. (2008). An integrated model of the recognition of *Candida albicans* by the innate immune system. *Nat. Rev. Microbiol.*2008;6:67-78.
  - 28- Williams DW., Kuriyama T., Silva S., Malic S., Lewis MA. *Candida* biofilms and oral candidosis: treatment and prevention. *Prionol.*2011;2000(55):250-265.
  - 29- Teodoro GR., Ellepola K., Seneviratne CJ., and Cristiane Y. Potential Use of Phenolic Acids as Anti-*Candida* Agents: A Review. *Front Microbiol.* 2015; 6: 1420.
  - 30- Faria NC., Kim JH., Goncalves L A., Martins M L., Chan K L., Campbell B.

- Enhanced activity of antifungal drugs using natural phenolics against yeast strains of *Candida* and *Cryptococcus*. *Lett. Appl. Microbiol.* 2011;52: 506–513.
- 31- Alves CT., Ferreira IC., Barros L., Silva S., Azeredo J., Henriques M. Antifungal activity of phenolic compounds identified in flowers from North Eastern Portugal against *Candida* species. *Future Microbiol.*2014; 9:139–146.
- 32- Wang SS., Wang DM., Pu WJ., Li DW. Phytochemical profiles, antioxidant and antimicrobial activities of three *Potentilla* species. *BMC Complement. Altern. Med.* 2013; 13:321
- 33- De Vita D., Friggeri L., D'auria FD., Pandolfi F., Piccoli F., Panella S., et al. Activity of caffeic acid derivatives against *Candida albicans* biofilm. *Bioorg. Med. Chem. Lett.*2014; 24:1502–1505.
- 34- Strugstad MP. And Despotovski SA. summary of extraction synthesis and potential. uses of juglon literature review. *Journal Of Ecosystem and Management* .2012; 13(3):1-16.
- 35- Paulsen MT. and Ljungma M. the natural toxin juglone causes degradation of p35 and include rapid H2AX phosphorylation and cell death in human fibroblast. *Toxicology and Applied Pharmacology.* 2005;209 (1): 1-9.
- 36- Inbaraj J. and Chignell CF. Cytotoxic Action of Juglone and Plumbagin: A Mechanistic Study Using HaCaT Keratinocytes. *Chemical Reasearch in Toxicology.* 2004; 17(1):55-62.



**Figure (1):**Type of extract (a) Water extract (b) Water-Methanol extract



**Error bars: 95% CI**

**Figure(2):** Mean of inhibition zones of candida albicans (mm) to different agents (Agar well diffusion technique)

Table (1) Mean and Standard Deviation(SD) for inhibition zones of Candida Albicans to different agents.

Concentration s	Mean(mm)			±SD		
	Extract			Extract		
	Water	Water-Methanol	Total	Water	Water-Methanol	Total
0.5%	9.46	11.40	10.43	.50	.50	.50
1%	10.46	12.40	11.43	1.00	1.00	1.00
5%	12.50	17.80	15.15	5.00	5.00	5.00
10%	12.70	18.36	15.53	10.00	10.00	10.00
15%	13.96	19.36	16.66	15.00	15.00	15.00
20%	15.10	21.06	18.08	20.00	20.00	20.00
25%	16.60	22.36	19.48	25.00	25.00	25.00
30%	17.60	23.76	20.68	30.00	30.00	30.00
40%	20.20	24.98	22.59	40.00	40.00	40.00
50%	22.10	25.84	23.97	50.00	50.00	50.00
<b>Total</b>	15.07	19.73	17.40	3.99	4.74	4.95
<b>Nystatin</b>	28.8			0.570		
<b>Amphotricin</b>	13.26			0.251		

No. of Isolates: 5

Table (2) Comparison between two types of Derum extracts according to Candida Albicans sensitivity.

Concentration	Group1	Group2	Sig.
0.5%	Water	Water-Methanol	0.157 (NS)
1%	Water	Water-Methanol	0.386 (NS)
5%	Water	Water-Methanol	0.001 (HS)
10%	Water	Water-Methanol	0.000 (HS)
15%	Water	Water-Methanol	0.000 (HS)
20%	Water	Water-Methanol	0.000 (HS)
25%	Water	Water-Methanol	0.000 (HS)
30%	Water	Water-Methanol	0.003 (S)
40%	Water	Water-Methanol	0.006 (S)
50%	Water	Water-Methanol	0.006 (S)

P&lt;0.05 Significant(S), P&lt;0.001 High significant(HS), P&gt;0.05 N0 significant(NS)



**Table (3)** Comparisons between different concentrations of Water and Water-Methanol extracts with Nystatin and Amphotricin B .

Group	Conc.	MD		Sig.	
		Nystatin	Amphotricin	Nystatin	Amphotricin B
Water	0.5%	-19.34	-3.80	.000(HS)	.008(S)
	1%	-18.34	-2.80	.000( HS)	.132(NS)
	5%	-16.30	-.76	.000( HS)	.921(NS)
	10%	-16.10	-.56	.000( HS)	.938(NS)
	15%	-14.84	.70	.000( HS)	.672(NS)
	20%	-13.70	1.84	.000( HS)	.068(NS)
	25%	-12.20	3.34	.000( HS)	.005(S)
	30%	-11.20	4.34	.000( HS)	.029(NS)
	40%	-8.60	6.94	.000( HS)	.001(HS)
	50%	-6.70	8.84	.000( HS)	.000( HS)
Water-Methanol	0.5%	-17.40	-1.86	.000( HS)	.065(NS)
	1%	-16.40	-.86	.000( HS)	.252(NS)
	5%	-11.00	4.54	.000( HS)	.000( HS)
	10%	-10.44	5.10	.000( HS)	.000( HS)
	15%	-9.44	6.10	.000( HS)	.000( HS)
	20%	-7.74	7.80	.000( HS)	.000( HS)
	25%	-6.44	9.10	.000( HS)	.000( HS)
	30%	-5.04	10.50	.000( HS)	.000( HS)
	40%	-3.82	11.72	.003(S)	.000( HS)
	50%	-2.96	12.58	.004(S)	.000( HS)
	Nystatin		15.54		.000( HS)

P<0.05 Significant(S), P<0.001 High significant(HS), P>0.05 NO significant(NS)