

Estimation of Erythromycin concentration In saliva of healthy volunteers

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Abstract

- Erythromycin is an extensively used group of antibiotic medicines and dentistry. Clinical pharmacology for erythromycin in saliva was not clear until the time of this research. The evaluation was achieved by using suitable method (efficient, low cost, and reproducible). The purpose of this study is the determination of the erythromycin concentration in saliva.
- **Methods:**Ten Subjects were given orally a single dose of 500 mg erythromycin very 6 hours. Samples 0.5 ml saliva was collected into centrifuge tubes at 0, 0.5, 1, 1.5, 2, 2.5, 3, 4, 6, 8, 12, and 24 hrs after dosing. Samples were centrifuged and the superannuated were injected to HPLC using USP-27 method of assays (2).
- **Results:**The results of this study indicated that erythromycin concentration in saliva ranged from 0.11-0.29 μ g. / mL. The other pharmacokinetic parameters are: the mean concentration of erythromycin C max 0.27±0.08 μ g /ml, AUC 0-∞ 217.25±9.25 μ g. h/ml, and Tmax. 7.28 ± 0.14 hr. and T *1*/2 8.33±2.68 hr.
- **Conclusion:** There was possibility to detection of erythromycin in saliva The HPLC method provided successful methods for monitoring the erythromycin with a detection limit reach 0.06 ng / ml. The analysis method is sensitive, reproducible, low cost and efficient for low concentration. The detection of erythromycin in saliva represents the distribution of the drug in saliva and indicated the suitability for erythromycin in treatments of dental and oral infections.
- Key word: bio-bioavailability of erythromycin concentration of erythromycin in saliva

Introduction

Erythromycin is a broad-spectrum macrolide antibiotic which possesses antimicrobial activity against grampositive and a few gram negative micro organisms. ⁽¹⁾ Its widely used in dentistry for treatment of oral infections such as pericoronitis, gingivitis, and chromatic conditions associated with secondary infections, especially for patients who are allergic

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to penicillin's. (2) Erythromycin is one of the drugs recommended by the American heart association for prevention of bacterial endocarditic in susceptible patients. ⁽³⁾ Erythromycin and its salts and esters are generally well tolerated and serious adverse Gastrointestinal effects are rare. disturbances such abdominal as cramp, discomfort and nausea, vomiting, and diarrhoea are fairly common after both oral and parenteral use, probably because of the stimulant activity of erythromycin on the gut. ⁽⁴⁾ Gastrointestinal effects are doserelated and appear to be more common in young than in older subjects. Suprainfection with resistant organisms may occur and pseudomembranous colitis has been reported. ⁽⁵⁾ All forms of erythromycin should be used with care in patients with existing liver disease or hepatic impairment, and the estolate is best avoided in such patients.⁽⁶⁾ Repeated courses of the estolate or use for longer than 10 days increase the risk of hepatotoxicity. Erythromycin may aggravate muscle weakness in patients with myasthenia gravis. Erythromycin and other macrolides have the potential to interact with a large number of drugs through their action on hepatic cytochrome P-450 isoenzymes, particularly CYP1A2 and CYP3A4. ^(9,10) Such interactions can adverse effects. result in severe including ventricular.⁽¹¹⁾

Materials and methods

Reagents and Solutions:

All chemical reagents used in this study were either from Fluka or BDH companies suitable for HPLC analysis. Erythromycin 250 mg caps were obtained from SDI Company⁽¹⁾

Weight variation. content uniformity, assay and dissolution studies were all carried out according to USP procedures ⁽¹²⁾.

Ten healthy Iraqi males were used, without any pathologic conditions. Their age ranged from 25-28 years. Each person was given 500 mg. of erythromycin caps every 6 hours for one day. Subjects were given orally a single dose of 500-mg tablets in a randomized fashion with 200 ml of water. Foods and drinks (other than water, which was allowed after 2 hours) were not allowed for 4 hours after dosing to all volunteers. Samples of approximately 0.5 ml saliva samples were collected into centrifuge tubes at (0 hr) and 0.5, 1, 1.5, 2, 2.5, 3, 4, 6, 8, 12, 24, 36 and 48 hr. after dosing. Participants were reminded for sample collection by cell phones calls. The centrifuge samples were centrifuged at 3000 rpm for 15 min; plasma samples were separated and kept frozen at -4 °C in coded glass tubes (12,13).

Chromatographic Conditions:

A reversed phase HPLC method was developed to quantities salivary levels of erythromycin. The chromatographic procedure may be carried out using a column (25 cm \times 4.6 mm) packed with stvrenedivinylbenzene copolymer (8 µm) with a pore size of 100 nm (PLRP-S is suitable), as mobile phase at a flow rate of 2.0 ml per minute a solution prepared in the following manner: to 50 ml of a 3.5% w/v solution of dipotassium hydrogen orthophosphate adjusted to pH 9.0 with 1Morthophosphoric acid add 400 ml of water, 165 ml of 2-methylpropan-2-ol and 30 ml of acetonitrile and dilute to 1000 ml with water and (c) a detection wavelength of 215 nm. Maintain the temperature of the column at 70° using a water bath for the column and at least one third of the tubing preceding the column. The flow rate is about 2 mL per minute ⁽¹²⁾.

Sample Preparation

300 µL of saliva from each sample was injected into a 5 ml test tube by using micropipette. For protein precipitation 100 µl of 10 % zinc sulfate was added, samples were vortexed, placed in refrigerator for 15 min and centrifuged at 3000 rpm for 15 min. Supernatant layer was separated of which 20 µL was injected onto the column and peak areas were recorded (14)

Results

The assays, dissolution as well as other USP investigations are listed in (table 1). The separation chromatogram of erythromycin is plotted in figure 1. The retention time of erythromycin was 5.88 min (Figure 1). This allows for the analysis of about 3 samples per h. the obtained chromatogram recorded broad peak of erythromycin. The retention time as well as the area under the curves is listed in (table 2). The method is highly sensitive, with the lower limit of quantitation of erythromycin at 0.05 The calibration curve was μg/ml. linear in saliva with a regression of The concentrations of r=0.99990. erythromycin in saliva were ranged from $0.11-0.29 \ \mu g/ml$ (table 3). Despite the fact that many modern HPLC methods have been described for the assay of erythromycin there is justification for investigating newer methods, especially where they are more cost-effective and there is an appreciable reduction in the procedural time compared to older methods.⁽¹²⁾ It may be applied to both research and therapeutic drug monitoring as demonstrated by the concentration time (pharmacokinetic curve in curves Figure 2). While the pharmaco-kinetic parameters are shown in (table 4). In conclusion, the HPLC assay method developed in this study using a reversed-phase system was found to be simple, rapid, not expensive and sensitive for assaying erythromycin concentration as low as 0.06 µg/ml in saliva.

Discussion

All products the met pharmacopoeia specifications for. variation, weight assay. and. dissolution profiles (table 1). The dissolution test revealed that $88.5 \pm$ 2.54 of stated erythromycin was released from SDI Iraq (n = 6). Therefore, either formulations met the USP dissolution specifications stating that not less than 80% of drug content should be released at dissolution time (12)

HPLC assay:

High performance liquid chromatography represent erythromycin can easily be detected after 5.88 minutes of capsules ingestion.

All chromatograms were free from any interference at the retention times of erythromycin, and both compounds were eluted completely and appeared as one big separate resolved peak with peak tailing followed with small one which could be related to the erythromycin isomers, it was possible to calculate peak height or peak area of standard curves. The retention times for erythromycin were 5.88 min. Linear relationships were found when the peak area ratios of erythromycin to the standard of erythromycin were plotted versus the erythromycin saliva concentration ranging from 0.11-0.29 $(r^2 = 0.99990).$ The use of external erythromycin standard to increase the accuracy of the assay whose availability is an important issue in HPLC assays and avoid interferences of other internal standard such as capsules additive.

Erythromycin was well detected in saliva of all volunteers. This method was very simple for monitoring of erythromycin in saliva. The short time of analysis, simplicity and sufficient makes sensitivity the method particularly useful for pharmacokinetic bioequivalent studies and of erythromycin even following oral single dose (1 caps/6 hrs). Based on estimated pharmacokinetic parameters and statistical analyses ⁽¹⁶⁾. These finding support our recommendations for using erythromycin in treatments of dental infections and oral soft tissues ^(17,19), practically, traumatic ulcers end even aphus ulcers associates with (20-22) infections secondary In summary, a rapid, sophisticated and sensitive HPLC method is described for determination of erythromycin in human saliva.

References

- 1- Nursing drugs handbook, 23 editions Lippincott W., pp: 138-139, 2003
- 2- Barry AL, et al. Macrolide resistance among Streptococcus pneumoniae and Streptococcus pyogenes isolated from outpatients in the USA. J Antimicrob Chemother 1997; 40: 139–40. PubMed
- Drici M-D, *et al.* Cardiac actions of erythromycin: influence of female sex. *JAMA* 1998; **280**: 1774–6.
- 4- Bingen E, *et al.* Resistance to macrolides in Streptococcus pyogenes in France in pediatric patients. *Antimicrob Agents Chemother* 2000; **44**: 1453–7.
- 5- Tateda K, *et al.* Direct evidence of antipseudomonal activity of macrolides: exposure-dependent bactericidal activity and inhibition of protein synthesis by erythromycin, clarithromycin, and azithromycin. *Antimicrob Agents Chemother* 1996; **40**: 2271–5.
- 6- Bui KQ, *et al.* In vitro and in vivo influence of adjunct clarithromycin on the treatment of mucoid Pseudomonas aeruginosa. *J Antimicrob Chemother* 2000; **45:** 57–62.

- 7- Ellsworth AJ, *et al.* Prospective comparison of patient tolerance to enteric-coated vs non-enteric-coated erythromycin. *J Fam Pract* 1990; 31: 265–70.
- 8- Seifert CF, *et al.* Intravenous erythromycin lactobionate-induced severe nausea and vomiting. *DICP Ann Pharmacother* 1989; **23:** 40–4.
- 9- Honein MA, *et al.* Infantile hypertrophic pyloric stenosis after pertussis prophylaxis with erythromycin: a case review and cohort study. *Lancet* 1999; **354:** 2101–5.
- 10- Mahon BE, *et al.* Maternal and infant use of erythromycin and other macrolide antibiotics as risk factors for infantile hypertrophic pyloric stenosis. *J Pediatr* 2001; **139**: 380–4
- 11- Westphal JF. Macrolide-induced clinically relevant drug interactions with cytochrome P-450A (CYP) 3A4: an update focused on clarithromycin, azithromycin and dirithromycin. *Br J Clin Pharmacol* 2000; **50:** 285–95. <u>PubMed</u>
- 12- the united states pharmacopoeia the united states formality , by author of the united states pharmacopoeia convention Inc., meeting at Washington , D.C. January , 1, 2004
- 13- FAO/WHO Specifications, identity and purity of some antibiotics. Journal of Pharmaceutical Experimental Therapy 1969, 207, 15-19.
- 14- FAO/WHO Evaluation of certain veterinary drug residues in food. thirty Sixth Report of the Joint FAO/WHO Export committee on food Additives. WHO Technical Report Series 1990, 799.
- 15- Reynolds JEF (ed). MARTINDALE. The Extra Pharmacopoeia, ed 30. London, The Pharmaceutical Press, 1993, p 1192.
- 16- Ellsworth AJ, *et al.* Prospective comparison of patient tolerance to entericcoated vs non-enteric-coated erythromycin. *J Fam Pract* 1990; **31:** 265–70
- 17- Seifert CF, *et al.* Intravenous erythromycin lactobionate-induced severe nausea and vomiting. *DICP Ann Pharmacother* 1989; **23:** 40–4
- 18- Honein MA, *et al.* Infantile hypertrophic pyloric stenosis after pertussis prophylaxis with erythromycin: a case review and cohort study. *Lancet* 1999; **354:** 2101–5. Correction. *ibid.* 2000; **355:** 758.
- 19- Mahon BE, *et al.* Maternal and infant use of erythromycin and other macrolide antibiotics as risk factors for infantile hypertrophic pyloric stenosis. *J Pediatr* 2001; **139**: 380–4

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- 20- eo CJ, *et al.* Erythromycin accelerates gastric emptying after pancreaticoduodenectomy: a prospective, randomized, placebo-controlled trial. *Ann Surg* 1993; **218:** 229–38
- 21- Simkiss DE, *et al.* Erythromycin in neonatal postoperative intestinal dysmotility. . Eady EA, *et al.* Effects of benzoyl peroxide and erythromycin alone

and in combination against antibioticsensitive and -resistant skin bacteria from acne patients. *Br J Dermatol* 1994; **131**: 331–6.

22- Strayer M, Ibrahim M. Dental treatment needs of homebound and nursing home patients. *Community Dent Oral Epidemiol* 1991;19:176-77.

No.	Test	No of tabs	result
1	Wt. variation	20	3.35 %
2.	Disintegration time	6	4.12 min.
3.	Specification	6	Comply with USP
4.	Dissolution	6	88%
5.	assay	6	104 %

 Table 1:
 The USP parameters of erythromycin capsules

Table 2: The chromatogram of erythromycin separation

Replicate	Peak area	Retention time (minutes)
1	26670420	5.92
2	26770554	5.93
3	25982575	5.89
Av.	26474516.333	

Table 3.The erythromycin concentration in saliva

Patients number	Concentration	(µg/ml)
1	0.19	
2	0.22	
3	0.17	
4	0.11	
5	0.14	
6	0.23	
7	0.29	
8	0.25	
9	0.27	
10	0.16	

			Table 4.
Parameters	(mean ± SD)	P- values	The pharmaco- kinetic
С <i>max</i> . µg. /ml	0.27. ±0.08	0.187	
AUC θ - ∞ (µg.h/ml)	217.25±9.25	0.782	parameters
T max.(hr.)	7.28±0.14	0.630	for erythromyc in .
T 1/2 (hr.)	8.33±2.68	0.937	



Figure 1 the erythromycin separation chromatogram in saliva.



Figure 2. Changes of erythromycin concentration with time.